Evaluation of chemical composition and antimicrobial activities of *Scrophularia striata* essential oil on dental caries pathogens

Shahram Dadelahi¹, Farnaz Yousefi², Parisa Emami Golmarz³, Elham Taheri^{4*}

- 1. Department of Oral and Maxillofacial Surgery, Faculty of Dentistry, Tabriz University of Medical Sciences, Tabriz, Iran
- 2. Department of Microbiology, Urmia Branch, Islamic Azad University, Urmia, Iran
- 3. Department of Genetics, Ahar Branch, Islamic Azad University, Ahar, Iran
- 4. Department of Pharmaceutical Biotechnology, Faculty of Pharmacy, Tabriz University of Medical Sciences, Tabriz, Iran

*Corresponding author:Tel: +98 9192855301 Fax:-

Address: Department of Pharmaceutical Biotechnology, Faculty of Pharmacy, Tabriz University of Medical

Sciences, Tabriz, Iran

E-mail: elham.taheri1366@gmail.com

Received; 20/04/2020 Revised; 2/06/2020 Accepted; 11/07/2020

Abstract

Introduction: Oral diseases are among the most important worldwide infectious diseases. Due to drug resistance and the side effects of chemical drugs, the use of herbal medicines has increased. *Scrophularia striata* (*S. striata*) is a herbal flowering plant that is used in microbial infections. Therefore, this study aimed to evaluate the antimicrobial activity of *S. striata* essential oil on dental carrier's pathogens.

Materials and Methods: In this study, *S. striata* essential oil was prepared and its antimicrobial activity was evaluated by disk diffusion, minimum inhibitory concentration (MIC), and minimum bactericidal concentration (MBC) methods on dental caries pathogens *Streptococcus mutans* (*S. mutans*), *Lactobacillus rhamnosus* (*L. rhamnosus*), *Actinomyces viscosus* (*A. viscosus*), and *Candida albicans* (*C. albicans*). Moreover, the chemical composition of *S. striata* essential oil was evaluated by gas chromatography-mass spectrometry (GC-MS) method.

Results: Our results showed that the most antibacterial activity of *S. striata* essential oil was related to *A. viscosus* (22.9 mm), *L. rhamnosus* (21.7 mm), and *S. mutans* (16.9 mm) essential oil showed a low antifungal activity against *C. albicans*. The dominant chemical composition of *S. striata* essential oil was terpens (39.8%).

Conclusion: In general, *S. striata* essential oil has an appropriate antibacterial activity against oral pathogens. Therefore, it can be use in pharmaceutical industry to produce antimicrobial agents against dental caries and oral infectious diseases.

Keywords: Dental caries Antimicrobial, Scrophularia striata, Essential oil

Introduction

On the report of the World Health Organization (WHO), oral diseases are regarded as a widespread and non-communicable disease (1). Different kinds of microorganisms have a role in causing oral

diseases like dental caries and periodontal disease (2). Dental caries is the most common oral disease in industrialized countries which affects 60-90% of children and the majority of adult and streptococci, lactobacilli, and Actinomyces are the main causes of dental caries (3). A variety of chemical compounds

Copyright © 2020 Journal of Basic Research in Medical Science. This is an open access article distributed under the terms of the Creative Commons Attribution 4.0 International License(https://creativecommons.org/licenses/by-nc/4.0/), which permits copy and redistribute the material, in any medium or format, provided that the original work is properly cited.

are commercially available to prevent and treat dental caries, but these cause changes in the oral microbiome and cause undesirable side effects such as diarrhea, vomiting, and discoloration of the teeth (4). The use of natural compounds for the prevention, control, and treatment of dental caries has been considered (5). The side effects of herbal medicine are considerably lower than chemical drugs and extreme use of antibiotics and chemical antibacterial compounds lead to antibiotic resistance (6). In recent years, the extract and essential oil of a large number of natural compounds have been used in cosmetic and hygienic industries dues to their antimicrobial and antioxidant properties (7). Scrophularia striata (S. striata) is a member of the Scrophulariaceae family and mostly used as a medicinal herb. Scrophulariaceae family are nearly 220 genera and 3000 species which grow in most parts of Iran, Turkey, and Azerbaijan (8). The member of this family has been shown many biological effects including antimicrobial, antiviral, and anti-inflammatory properties (9). The S. striata contain cinnamic acid, flavonoids (quercetin), isorhamnetin-3-O-rutinoside, nepitrin, and phenylpropanoid glycoside which found in various parts of the plant (10). The various forms of S. striata have been used for the treatment of allergies, rheumatics. and chronic inflammatory diseases for a long time. Its extract can be used as an antibacterial against and has a role in decreasing edema, cell infiltration, and proliferation of activated T-lymphocytes in joint tissues (11). Also, it acts as an inhibitor of some inflammatory factors, and several shown that their antistudies have inflammatory properties are due to the presence of iridoids and phenylpropanoids (12, 13).

According to the antimicrobial properties of *S. striata*, it can be effective in the treatment of oral infections. Therefore, this study aimed to investigate the antimicrobial effects of *S.*

striata essential oil on dental carrier's pathogens Streptococcus mutans (S. mutans), Lactobacillus rhamnosus (L. rhamnosus), Actinomyces viscosus (A. viscosus), and Candida albicans (C. albicans) and evaluate its chemical composition.

Materials and Methods

Preparation of Essential Oil

The *S. striata* was collected from medical plants centers in Tabriz city and identified and approved bythe Herbarium of the Islamic Azad University, Tabriz Branch. The 100 gr *S. striata* was dried in a dark place and then powdered. The obtained powder was added to a balloon containing 600 ml of distilled water. The essential oil was obtained using a Clevenger apparatus (Zarin Pyrex, Iran) and sterilized using a 0.4 µg syringe filter. The prepared essential oil was stored at 4°C until required.

Preparation of Bacterial Strains

The studies dental caries pathogens in the present study were include *S. mutans*, *L. rhamnosus*, *A. viscosus*, and *C. albicans*. These established terminology bacterial strains were purchased from the Iranian Biological Resource Center-Persian Type Culture Collection (IBRC-PTCC).

Evaluation of Antibacterial Activity

Agar Disk Diffusion: The agar disk diffusion method is used to evaluate the antibacterial activity of *S. striata* essential oil. First, the standard 0.5 McFarland microbial suspension was prepared and cultured on blood agar medium (Merck, Germany). The antimicrobial susceptibility disks containing different concentrations of *S. striata* essential oil (50%, 25%, 12.5%, 6.25%, 3.12%, 1.56%) were used. The antibiotic disks include florfenicol (30 μg), enrofloxacin (5 μg), amoxicillin (25 μg), and penicillin (6 μg)

were considered as positive control and a disk containing the solvent of essential oil (distilled water) were used as a negative control. In the end, the inhibition zone diameter was assessed after 48 hours' incubation at 37°C.

Broth Micro-Dilution: The antibacterial activity of S. striata essential oil was also evaluated using broth microdilution method. The different concentrations of S. striata essential oil (50%, 25%, 12.5%, 6.25%, 3.12%, and 1.56%) were prepared using sterile Brain Heart Infusion (BHI) medium (Merck, Germany) in sterile tubes. The microbial suspensions with standard 0.5 McFarland concentration were prepared and then cultured in the BHI medium. The cultured bacterial strains without S. striata essential oil was considered as positive controls and BHI medium was considered as negative controls. The prepared tubes were incubated at 37°C for 24 hours and the least concentration of S. striata essential oil without opacity was considered as the Minimum Inhibitory Concentration (MIC). After treated strains were cultured on blood agar medium, the minimum concentration without bacterial growth was assumed as the Minimum **Bactericidal** Concentration (MBC).

Evaluation of Essential Oil Compounds

The gas chromatograph (Shimadzu-QP2010, Japan) with the ZB-WAX column (length 20 m, inner diameter 0.18 mm, thickness 18.1 um) was used to identify the compounds of the S. striata essential oil. The essential oil of S. striata was diluted with normal hexane and was injected into μl gas chromatography/mass spectrometry (GC/MS). The initial temperature of the oven was 50°C, maintained at this temperature for 5 minutes (thermal gradient: 3°C per minute) and then the temperature was increased to 240°C. The final temperature of the oven was 300°C and maintained at this temperature for

3 minutes (thermal gradient: 3°C per minute). The temperature of the injector was 300°C and split/splitless (1 to 50). Helium (99.9999%) was used as the carrier gas at a flow rate of 1ml/min. Then, spectrometry (Agilent 5973, USA) (length 20 m, inner diameter 0.25 µm, thickness 0.25 mm) was used. The temperature of the chamber was 150°C. ionization temperature of the detector was 230°C, the ionization energy was 70 eV, and the mass analyzer was Quadrupole. The scan mass range was 40 m/z to 550 m/z. The mass spectrometry was used to determine the compounds of the essential oil of S. striata. The spectral values were compared with Kovatz index values in the standard tables and the compounds of the essential oil of S. striata were identified according to data and information available in the GC-MS library.

Results

Antibacterial Activity

Agar Disk Diffusion: The obtained results showed that the largest growth inhibition zone was related to the *A. viscosus* (22.9 mm), *L. rhamnosus* (21.7 mm), and *S. mutans* (16.9 mm). Moreover, the growth inhibition zone of *C. albicans* was 5.1 mm. The *S. striata* essential oil showed a larger growth inhibition zone than the penicillin, enrofloxacin, florfenicol. The Amoxicillin created the largest inhibition zone in the studied bacterial strains (Table 1).

Broth Micro-dilution: The obtained results showed that the *A. viscosus*, *L. rhamnosus*, and *S. mutans* showed a high sensitivity (MIC=1.56% and MBC=3.12%) to *S. striata* essential oil. Also, the *C. albicans* showed a high resistance to *S. striata* essential oil.

Chemical Composition

According to the obtained results, 26 compound were identified in the *S. striata* essential oil, which was 88.4% of the

essential oil. The dominant chemical composition found in the *S. striata* essential

oil was terpens (39.8%) (Table 2).

Table 1. Inhibition zone diameter of *S. striata* essential oil on dental caries pathogens.

Bacterial strains	Inhibition zone diameter (mm)								
	Essential oil	Penicillin	Enrofloxacin	Amoxicillin	Florfenicol				
A. viscosus	22.9	19.9	19.2	24.3	18.7				
L. rhamnosus	21.7	17.8	19.0	18.8	16.6				
S. mutans	16.9	16.9	17.1	18.5	16.4				
C. albicans	5.1	2.7	5.4	3.8	3.5				

Table 2. The obtained compounds of *S. striata* essential oil using GC/MS.

No.	Compounds	Frequency	No.	Compounds	Frequency
1	Terpens	39.8	14	Tetradecanedioic	0.8
2	p-Cymene	14.5	15	α-Thujene	0.8
3	Palmitic acid	9.4	16	α-Phellandrene	0.7
4	Saturated fatty	6.7	17	β-Myrcene	0.7
5	Thymol	1.7	18	α-Pinene	0.6
6	Carvacrol	1.2	19	2-Decanol	0.5
7	Linalool	1.6	20	Benzyl benzoate	0.5
8	B-caryophyllene	1.6	21	trans-Nerolidol	0.5
9	β-Elemene	1.3	22	Phytol	0.5
10	p-Xylene	1.1	23	Isopulegol	0.4
11	2-Undecanone	1.0	24	Z-β-Damascenone	0.3
12	δ-Terpinene	0.9	25	Camphene	0.3
13	α-Terpineol	0.8	26	E,Z-Farnesol	0.2

Discussion

Recently, secondary metabolites of medicinal plants have been studied for antimicrobial effects (14), and it has been reported that most of the herbs have antifungal, antiphrastic, antibacterial and antiviral properties (15). Therefore, plant extracts have been widely used in the pharmacology, herbal pharmacology, microbiology, medical and clinical phytopathology and food preservation (16). Traditional herbal medicine has been used for treatment of various diseases for several centuries in many parts of the world, and these antibacterial agents have revolutionized the treatment of various bacterial infections (17). In this study, antibacterial and antifungal activity of S. striata essential oil on the most important oral pathogens such as S. mutans, L. rhamnosus and A. viscosus as well as C. albicans fungi were investigated by agar disk diffusion and broth microdilution methods.

The obtained results showed that the largest inhibition zone was related to A. viscosus (22.9 mm diameter) and C. albicans (5.1 mm diameter). In addition, the S. striata essential oil created a larger inhibition zone than the Enrofloxacin, Penicillin, and Florfenicol antibiotic in the studied strains. Many studies have investigated the antimicrobial activity of medicinal plants on various gram-negative and gram-positive bacteria and fungi. In a study by Safavi et al. reported that the largest diameter of inhibition zone (12 mm) due to S. striata essential oil was related to gram positive S. aureus (18). In another study by Moori Bakhtiari et al. reported that the largest diameter of inhibition zone (26 mm) due to S. striata essential oil related to S. aureus (19). According to the obtained results in present study and also mentioned studies it can be said that the essential oil of S. striata has an appropriate antibacterial activity against Gram positive and Gram negative bacteria as well as pathogenic fungi.

The obtained results from GC-MS assay showed that the dominant chemical composition of S. striata essential oil was terpens (39.8%). Previous studies reported that terpens has an antimicrobial activity on various pathogenic bacterial strains. The presence of this compound phytochemicals of S. striata essential oil can be a reason for the inhibitory potency of this essential oil on different bacterial strains (20). The presence of terpens in the phytochemicals of the essential oils of various medicinal plants can cause its antimicrobial activity against different bacterial strains (21). Phytochemical studies on essential oils of medicinal plants indicate the presence of terpens, and its antimicrobial activity indicates their inhibitory effect on Gram-positive and Gram-negative bacteria (22). Therefore, it can be said that this

References

- 1. Salehi B, Kregiel D, Mahady G, Sharifi-Rad J, Martins N, Rodrigues CF. Management of Streptococcus mutans-Candida spp. Oral Biofilms' Infections: Paving the Way for Effective Clinical Interventions. J Clin Med. 2020;9(2):517. doi: 10.3390/jcm9020517.
- 2. Yari Z, Mahdavi S, Khayati S, Ghorbani R, Isazadeh A. Evaluation of antibiotic resistance patterns in Staphylococcus aureus isolates collected from urinary tract infections in women referred to Shahid Beheshti educational and therapeutic center in Maragheh city. Med J Tabriz Uni Med Sci. 2020;41(6):106-12.
- 3. Petersen PE, Lennon MA. Effective use of fluorides for the prevention of dental caries in the 21st century: the WHO approach. Community Dent Oral Epidemiol. 2004;32(5):319-21.

inhibitory activity can be involved with this chemical compound.

Conclusion

The results of this study showed that the essential oil of *S. striata* can have inhibitory effects on bacterial strains (*S. mutans*, *A. viscosus* and *L. rhamnosus*) and fungi (*C. albicans*). Therefore, according to the herbal and native origin of this drug, and less side effects in compared to other chemical compounds and antibiotic, it can be used in pharmaceutical industry to production of antibacterial, disinfectants and mouthwashes drugs to control of infectious diseases and dental caries.

Aknowledgments

We thank the whole staff of Biotechnology Research Center, Islamic Azad University, Tabriz Branch, for assistance in the successful strategy of this research.

- 4. Mahdavi S, Azizi Dehbokri M, Isazadeh A. Contamination of chicken meat with salmonella spp distributed in mahabad city, iran. Int J Enteric Pathog. 2018;6(3):65-8. doi: 10.15171/ijep.2018.18.
- 5. Mahdavi S, Kheyrollahi M, Sheikhloei H, Isazadeh A. Antibacterial and Antioxidant Activities of Essential Oil on Food Borne Bacteria. Open Microbiol J. 2019;13(1):81-85.
- 6. Mahdavi S, Chalabi P, Zomorodi S, Isazadeh A. Effect of Bananas Puree on Survival of Lactobacillus casei in Coktel Apple and Banana Juice During Storage. Pharm Biomed Res. 2018;4(2):23-7.
- 7. Mahdavi S, Hazimian S, Isazadeh A, Babashpour M, Shishehgar R. Study of the antioxidant and antimicrobial effects of the ethanolic extract of Eucalyptus camaldulensis Dehnh against infectious bacteria isolated from clinical and animal

- sources. J Comp Pathobiol. 2017;13(4):2063-70.
- 8. Alaee M, Akbari A, Karami H, Salemi Z, Amri J, Panahi M. Antidiabetic and protective effects of Scrophularia striata ethanolic extract on diabetic nephropathy via suppression of RAGE and S100A8 expression in kidney tissues streptozotocin-induced diabetic rats. J Basic Clin Physiol Pharmacol. 2020;31(2). doi: 10.1515/jbcpp-2019-0186.
- 9. Singab AN, Youssef FS, Ashour ML, Wink M. The genus E remophila (S crophulariaceae): an ethnobotanical, biological and phytochemical review. J Pharm Pharmacol. 2013;65(9):1239-79. doi: 10.1111/jphp.12092.
- 10. Kerdar T, Moradkhani S, Dastan D. Phytochemical and biological studies of scrophularia striata from Ilam. Jundishapur J Nat Pharm Prod. 2018;13(3):1-5. doi: 10.5812/jjnpp.62705.
- 11. Mahboubi M, Kazempour N, Nazar AR. Total phenolic, total flavonoids, antioxidant and antimicrobial activities of striata Scrophularia Boiss extracts. Jundishapur J Pharm Nat Prod. 2013;8(1):15-9.
- 12. Rostami F, Ghasemi HA, Taherpour K. Effect of Scrophularia striata and Ferulago angulata, as alternatives to virginiamycin, on growth performance, intestinal microbial population, immune response, and blood constituents of broiler chickens. Poult Sci. 2015;94(9):2202-9. doi: 10.3382/ps/pev198.
- 13. Zengin G, Stefanucci A, Rodrigues MJ, Mollica A, Custodio L, Aumeeruddy MZ, et al. Scrophularia lucida L. as a valuable source of bioactive compounds for pharmaceutical applications: In vitro antioxidant, anti-inflammatory, enzyme inhibitory properties, in silico studies,

- and HPLC profiles. J Pharm Biomed Anal. 2019;162:225-33. doi: 10.1016/j.jpba.2018.09.035.
- 14. Mahdavi S, Isazadeh A. Lactobacillus casei suppresses hfq gene expression in Escherichia coli O157:H7. Br J Biomed Sci. 2019;76(2):92-4. doi: 10.1080/09674845.2019.1567903.
- 15. Taheri E, Ghorbani S, Safi M, Sani NS, Amoodizaj FF, Hajazimian S, Heidari M, Isazadeh A, Heidari M. Inhibition of Colorectal Cancer Cell Line CaCo-2 by Essential Oil of Eucalyptus camaldulensis Through Induction of Apoptosis. Acta Med Iran. 2020;58(6):260-5.
- 16. Mahdavi S, Isazadeh AR. Investigation of contamination rate and determination of pattern of antibiotic resistance in coagulase positive staphylococcus aureus isolated from domestic cheeses in Maragheh, Iran. Pathobiol Res. 2019;22(2):85-59.
- 17. Firouzi Amoodizaj F, Baghaeifar S, Taheri E, Farhoudi Sefidan Jadid M, Safi M, Seyyed Sani N, et al. Enhanced anticancer potency of doxorubicin in combination with curcumin in gastric adenocarcinoma. J Biochem Mol Toxicol. 2020;e22486. doi: 10.1002/jbt.22486.
- 18. Safavi F, Ebrahimi P, Mighani H. In vitro anti-bacterial activity of root and aerial parts of Scrophularia striata Bioss on Escherichia coli, Staphylococcus aureus and Bacillus cereus. Armaghane Danesh. 2013;18(8):603-14.
- 19. Moori Bakhtiari N, Jowzi L. In vitro Effect of Aqueous and Ethanolic Extracts of Scrophularia striata on Some Respiratory and Urinary Bacterial Pathogens. J Ardabil Univ Med Sci. 2016;15(4):423-31.
- Kerdar T, Moradkhani S, Dastan D. Phytochemical and biological studies of scrophularia striata from Ilam.

- Jundishapur J Nat Pharm Prod. 2018;13(3):e62705. doi: 10.5812/jjnpp.62705.
- 21. Nikkhah E, Asnaashari S, Babaei H, Heshmati Afshar F, Delazar A. Chemical composition and biological activities of essential oil and methanol extract of
- Scrophularia umbrosa. Res J Pharmacognosy. 2017;4(1):41-50.
- 22. Amiri H, Lari YH, Esmaeili A, Samsamnia F, Eghbali D, Viskarami GH, Dosti B, Noormohamadi E. Essential oil composition and anatomical study of Scrophularia striata Boiss. Iran J Med Aromatic Plant Res. 2011;27(2):271-278.